Chemical Systematics of the Verbenaceae

## CHAPTER 3

## CHEMICAL SYSTEMATICS OF THE VERBENACEAE

## 3:1 Introduction

The Verbenaceae, a large family of 75 genera and 2500 species (Jafri and Ghafoor, 1974) is chiefly tropical and subtropical in distribution, although a few species extend into temperate regions of the new world.

Economically the family is important for teak (<u>Tectona grandis</u>), which is highly prized for its wood. A number of genera contain important ornamentals, notable among them are, <u>Clerodendrum</u>, <u>Duranta</u>, <u>Holmskioldia</u>, <u>Lantana</u>, <u>Petrea</u>, <u>Verbena</u> and <u>Vitex</u>.

In India the family is widely distributed upto an altitude of 4,000 feet (Clarke, 1885). However, most of its members are confined to central India. Clerodendrum, Verbena etc. are introduced from temperate regions of the world.

Avicennia is confined to coastal regions of India.

Most of the Verbenaceae members are predominantly scandent or straggling shrubs, rarely herbs (<u>Verbena</u>), armed shrubs with axillary thorns (<u>Duranta</u>) or deciduous trees (Tectona) or Halophytes as in case of Avicennia. The leaves

are simple, opposite deccusate, exstipulate; except in <a href="Vitex">Vitex</a> which show compound leaves. The presence of a pair of glands at the base of lamina is a characteristic feature of <a href="Citharexylum">Citharexylum</a>. Inflorescence is variable, usually determinate and one or other modification of dichosial cyme. Racemes with flowers arranged in dense or lax spike or corymbs are seen in <a href="Verbena">Verbena</a>, whereas the inflorescence is paniculate in <a href="Gmelina">Gmelina</a> philippensis and <a href="Tectona grandis">Tectona grandis</a>.

Flowers are typically bisexual, hypogynous and pentamerous. The calyx shows considerable variation in number of lobes or teeth. It is usually cupular having 5 lobes or teeth and persistent with the fruit. In Phyla the calyx is generally 2-4-dented, whereas 5 lobes and 5 scales in the throat are seen in Petrea. Calyx is membranous in Lantana. In Tectona the calyx is campanulate and 5-7-lobed. In other members like Vitex, Clerodendrum, and Holmskioldia the calyx is perfectly 5-toothed and slightly enlarged in fruit. The gamopetalous corolla is cylindrical or infundibuliform with as many lobes as the calyx. The lobes are mostly unequal but at times 5 equal or subequal lobes are present in genera like Gmelina, Holmskioldia and Premna. Hypocrateriform corolla with 5-7 subequal lobes is present in Tectona.

In the family, the stamens are invariably 4 and didynamous, inserted on the corolla tube and alternating with corolla lobes. The fifth stamen is generally represented by a staminode. Tectona is the only exceptional taxon showing 5 fertile stamens. The anthers are dithecous and mostly dehisce longitudinally. In Stachytarpheta, of the 4 stamens only the 2 anterior ones are fertile and possess divergent anther lobes. Anther lobes sagittate in Duranta. Normally the stamens are included but exceptionally exerted in Tectona, Clerodendrum and Holmskioldia.

The gynoecium is usually bicarpellary; some times it is 4-carpelled as in <u>Duranta</u>. Ovary is superior, 4-lobed, 2-4-loculed with one ovule in each apparent cell on axile placenta. Ovules usually solitary, mostly erect, anatropous with micropyle facing downward. The capitate stigma is usually bifid but with unequal 4 lobes as in <u>Duranta</u>. A curved and acutely bifid stigma occurs in <u>Clerodendrum</u>. Fruit is generally a drupe with as many pyrenes as ovules in ovary or nutlets (<u>Verbena</u>). The fruit is a 2-4-valved capsule in <u>Avicennia</u>. The seeds are with a straight embryo and exalbuminous. However, fleshy endosperm is present in Avicennia, Stilbe, Chloanthes.

# 3:2 Taxonomy

The family Verbenaceae was included under subclass

Gamopetalae, series Bicarpellatae and order Lamiales by

Bentham and Hooker (1876). Engler and Prantl

(1897) grouped it close to Lamiaceae under suborder

Verbenineae of the order Tubiflorae. This scheme was later

followed by Engler and Diels (1936). In the latest edition

of Engler's syllabus der pflanzenfamilien (ed. Melchior,

1964), the earlier classification is maintained. Order

Tubiflorae encompasses Verbenaceae under "Unterreiche"

Verbenineae.

Hallier (1912) recognised 4 primary sub divisions for the various families of the Dicotyledons. The IVth sub division 'ochnigenes' comprised, amongst other families, Verbenaceae under the order Tubiflorae. Bessey (1915) on the basis of corolla zygomorphy and gynoecium character put Verbenaceae and Lamiaceae in a distinct order, the Lamiales. The order Lamiales according to him consists of Verbenaceae, Myoporaceae, Phrymaceae and Lamiaceae (Labiatae). Rendle (1925) and Wettstein (1935) followed the concept of Engler and Prantl (1897) regarding the placement of the Verbenaceae.

Hutchinson (1926) treated Verbenaceae and its allied family Phrymaceae under Lamiales, but latter (1948)

segregated them to form order Verbenales, a group that represented the climax among the woody dicots. In Verbenaceae, if one looks for a primitive type, the choice falls on the genus <u>Tectona</u>, a hard wooded timber tree in which 5 stamens have been retained. In all other genera, the 5th stamen is absent or rarely represented by a staminode. Therefore, Hutchinson considered this group as unrelated to Labiatae and derived it from rubiaceous stock. Thus Verbenaceae was regarded as parallel to and not as a close relative of Lamiales. Hence, it topped the linear sequence of Lignosae while the Lemiaceae topped the Herbaceae. In the latest edition of Families of Flowering Plants (1973) he recognised as many as 5 families in order Verbenales namely Ehretiaceae, Verbenaceae, Stilbaceae, Dichrostyladaceae (Chloanthaceae) and Phrymataceae.

Skottsberg (1940) grouped Verbenaceae under Tubiflorae, while Gundersen (1950) erected a new order Boraginales wherein the Verbenaceae was grouped along with families such as Boraginaceae, Myoporaceae, Labiatae and Callitrichaceae. Lawrence (1951) adopted the system proposed by Engler and Diels (1936) and treated Verbenaceae under Tubiflorae. Benson (1957) treated this family under separate order Lamiales. This was also supported by Soo (1961). The systems proposed by Cronquist (1968) and Takhtajan (1969,1980)

Verbenaceae under Lamiales in sub class - Asteridae of the Dicotyledons (Magnoliatae). The families included under Lamiales are Labiatae, Boraginaceae, Phrymaceae, and Callitrichaceae. Cronquist (1968) visualises a close relationship of the Verbenaceae to the Labiatae. To quote him "Indeed it is difficult to know where to draw the line between them, and a case might be made for treating the present Verbenaceae and Labiatae as the more primitive and more advanced segments of a single family".

The family Verbenaceae is classified into 8 tribes by Bentham and Hooker (1876) on the basis of inflorescence character, number of locules in the ovary and nature of ovule. The various tribes recognised are Phrymeae, Stilbeae, Chloantheae, Verbeneae, Viticeae, Caryopterideae, Symphoremeae and Avicenniae (Table 11).

Briquet (1897) based the infrafamilial or tribal classification on cohesion of ovary and number of ovules in each locule. The family is sub-divided into 8 subfamilies and 15 tribes. This treatment is followed in the latest edition of Engler's syllabus (ed. Melchior, 1964). Hutchinson (1969) elevated Bentham and Hooker's tribes Stilbeae, Chloantheae and Phrymeae to separate familial rank. Thus the family Verbenaceae as now circumscribed

VERBENACEAS, taxa screened and their arrangement in - Table-;=71.

values schemes of classification

HUTCHINSON - MOLDENKE (1926), (1969) (1959)	FAMILY PHRYNACEAE ZAMILY PHRYMACEAE	STILBACEAE STILBACEAE	CHLOATTHACEAE SUBFLATIY CHLOANTHOLDEAL	SE VIRE ENERS SUBPACILIX VANSENOIDE AR							TRIBE <u>VITICEAE</u> SUBPATILY <u>VITICOLDEAE</u> E	TRIBE CARYOPTERIDERS SUSFAMILY CARYOPTERIOCIDE-E	TRIBE SYMPHOREMEAE PAMILY SYMPHOREMACERE	TRIBE AVICEMMIRAE FAMILY AVICEMMIRCHE	
BRIQUET HU (192	FAUI	DIV.B SUBFAMILY STILBOIDEAE	Div. A SUBE. ILY CHIO. NITHOLDES	SUBFAMILY TREEDMOIDEAE	TRIBE JANTANEAE	THIBE VERBENEAE	TRIBE PATRACEASE	TRIBE <u>CITHAREXYLEAE</u>	,	VITAL TRIBE TECTONIAE	뛲	SJERWILY CARYOPTURIDOIDEAE	SUBFAIILY SYMPHOREMATOIDEAE	SUBFAILLY AVICENIOLIERS TRI	SUBEATIN NYCHANGRAER
B.E.N.T.HAM and HOOKER (1876)	TRIBE I PERTMEAE	II STILBEAE	TII CHLOANTHEAE	IV VERBUNEAS	<u>Lantana</u> <u>phyle</u> Stachytaroheta	Verbena	Petres	Citharexylum }	V VITICEAE	Tectona	Clerodendrum, Colmskioldia	VI CARYOPTERIDEALE Dav. B	VII SYAFHORBIJAE Drv.C	VIII AVICEMNIE E	Δ 1. 1. 1. 1. 1. 1. 1. 1. 1. 1. 1. 1. 1.

includes only tribes Verbeneae, Viticeae, Caryopterideae, Symphoremeae and Avicenniae.

Many controvercies exist regarding the position of the genus Avicennia. Endlicher (1838) in his classification of Verbenaceae, appended Avicennia as an unnumbered tribe Avicenniae. Latter in 1843, he proposed separate monogeneric family Avicenniaceae. Schauer (1847), Bentham and Hooker (1876) however, maintained Avicennia as constituting a separate and last tribe of the Verbenaceae.

Avicennia, the status of a sub-family Avicennioideae Briq.

Moldenke and Moldenke (1946) and Moldenke (1959) accepted

Avicenniaceae of Endlicher as a family distinct from

Verbenaceae under the Lamiales, on the basis of wood anatomy,

articulate branches, free central placentation and pendent

orthotropous ovule. Moldenke (1959) also recognized

Phrymaceae, Stilbaceae and Symphoremaceae as separate

families.

#### 3:3 Earlier chemical work

The known chemical data (Table 12) of various

Verbenaceae taxa include flavonoids, lignans, quinones,

alkaloids and steroidal saponins. The phenolic chemistry is

Callicarpagnacrophylla Lvs. 5,41- Callicarpagnacrophylla Lvs. 6,41- Callicarpagnacrophylla Lvs. Hispin Callicarpagnacrophylla Lvs. Hispin Callicarpagnacrophylla Lvs. Hispin Callicarpagnacrophylla Lvs. Hispin Callicarpagnacrophylla Lvs. 7-c g C. incrifolium Lvs. Scute C. serratum C. trichotomum C. trichotomum Lvs. Scute C. serratum C. trichotomum Lvs. Acace C. serratum C. trichotomum Lvs. Scute Scute C. aspatica Lvs. Acace Cuarp Lvs. Acace Cuarp Cuarp Casalatica Lvs. Acace Cuarp Cuarp Cuarp Casalatica	Eart Compound reported  Lys. 5,4'-di-0F,5;7;3'-tri-Ome flavone  Lys. Hispidian, Soutellaran  1.9. Hispidian, Soutellaran  1.9. Dut no free aglyrones.  Als. Apigenin, Acacetin, Acacetin  Lys. Soutellare: 1-4'-arabinoside, soutellarein  Lys. Soutellare: 1-4'-arabinoside, soutellarein  Lys. Soutellare: 1-4'-arabinoside, soutellarein  Lys. Soutellare: 1-4'-arabinoside, soutellarein  Lys. Soutellarein, Acacetin  Lys. Acacetin-7-p-glucoso (1-2)p-glucuronide  Lys. Mclyndin  Soutellarein, Pectolinoringenin  Lys. Hispidian, Setolinoringenin  Lys. Hispidian, Setolinoringenin  Lys. Lutsolin  Lys. Mclyndin  Soutellarein, Setolinoringenin  Lys. Lutsolin  Lys. Lutsolin  Apigenin, Lutsolin, Amercetin.  Glucuronide, Api-7-glucuronide, Lutso-1-  glucuronide, Api-7-glucuronide, Lutso-7-  glucuronide, Cerotenoide, Lutso-7-  glucuronide, Cerotenoide, Lutso-7-  glucuronide, Cerotenoide, Lutso-7-  glucuronide, Cerotenoide, Lutso-7-	;;;;;;;;;;;;;;;;;;;;;;;;;;;;;;;;;;;;;;			<b>9</b>
Callicarder name  Part Compound reported  (Naviords State Lara )  Callicarder control of the con	C. Internet and subservations also supposed to the control of the	١		EARLIN CHENICAL WORK	
Callicangementocohylla  Callicangementocohylla  Callicangementocohylla  Callicangementocohylla  Callicangementocohylla  Ive. H.sp.dullan, Scutellaran  C. incrime  Ive. Soute-laran, careetan  Ive. Acacetan, firstdallan  Soute-laran, Pertolinoringenin  Ive. Inteolin  C. introlin  Acacetan, introlin, introlin,  C. introlin  Acacetan, introlin, introlin,  C. introlin  Acacetan, introlin, introlin,  Acacetan, introcade, Api-7-glucuronide, Iuteo-7-  Religion  C. introline  Acacetan, introlin, introlin,  Acacetan, introlin,  Acacetan, introline  C. introline  C. introline  Acacetan, introline  C. introline  C. introline  Acacetan, introline  C. introline  C. introline  Acacetan, introline  C. introline  Acacetan, introline  Acacetan, introline  C. introline  Acacetan, introline  C. introline  Acacetan, introline  Acacetan, introline  Acacetan, introline  Acacetan, introline  Acacetan, introline  Acacetan, introlin	Callegrage accoonding  Callegrage according  Callegrage according  Callegrage  Callegr		part		Author
CallicarDespectrophylla  1.vs. 5,4'-dn-0F,3,7,3'-tr1-0me flavone Citherexylum subservatum  1.vs. Hayndhin, Suvelaharan  1.vs. Hayndhin, Soutelaran  2. infortumetum  2. infortumetum  2. infortumetum  3.13. Angenun, Acecetun  1.vs. Soutellaran, Acecetun  2. erretum  2. tr1-chotomum  3. soutellaran, Acecetun  4.vs. Soutellaran, Acecetun  5. soutellaran, Englandin  6. off intending arborne  6. off intending arborne  1.vs. Acecetun-7-p-glucoso (1-2)p-glucuronide  6. off intendin, Caffete and Ferulic acid.  6. off intendin, Caffete and Ferulic acid.  7. soutellaran, Latentum  1.vs. Acecetun-7-p-glucoso (1-2)p-glucuronide  6. off intendin, Caffete and Ferulic acid.  7. soutellaran, Pertolinoringenin  1.vs. Acecetun-7-p-glucoso (1-2)p-glucuronide  6. off intendin, Caffete and Ferulic acid.  7. curretage  8. cutellaran, Pertolinoringenin  1.vs. Acecetun-7-p-glucoso (1-2)p-glucuronide  6. off intendin, Latentum  1.vs. Acecetun-7-p-glucoso  1.vs. Intendin  1.vs. Intendin  2. experice acectun  3. cutellaran, Pertolinoringenin  1.vs. Intendin  1.vs. Intendin  2. experice acectun  3. cutellaran  4. casectun  6. aspetica  6. aspetica  7. cutellaran  7. cutellaran  8. cutellaran  1.vs. Intendin  1.vs. Intendin  1.vs. Intendin  1.vs. Intendin  1.vs. Intendin  1.vs. Intendin  1.vs. Acecetun  2. experice acectun  4. casectun  4. casectun  6. aspetica  6. aspetica  7. cutellaran  8. cutellaran  1.vs. Intendin  1.vs. In	Callicarps mecrophyla  1.75, 7.47-dn-0F,57,73-tr1-0m flavone Citarestium subservetum  1.75, 1.10m-2-5,41-tr1-0M flavone Clascaderum indicum  1.75, 1.10m-2-5,41-tr1-0M flavone  1.75, 1.10m-2-5,41-tr1-0M flavone  1.75, 1.10m-2-5,41-tr1-0M flavone  2. infortung tum  3. infortung tum  4. infortung tum  5. infortung tum  6. infortung tum  7. infortung tum  6. infortung tum  7. infortung tum  8. infortung tum  8. infortung tum  1. infortung  1. infortung tum  8. infortung tum  8. infortung tum  8. infortung  1. infortu	FLAVONOTDS		-	
Citroreoxilum gubserreitum  Lvs. Hespilulin, Suctalarean  Clarodendrum Andicum  1'9. Otherwoodee of Rispidulin, Soutellarean  C. inerme  Lvs. Scutellarean, Acacetin  Lvs. Scutellarean, Acacetin  Lvs. Scutellarean expension  C. inerme  C. inerme  Lvs. Scutellarean, Pertolinoringenin  Lvs. Scutellarean expension  C. inerme  C. inerme  C. inerme  C. inerme  Lvs. Scutellarean, Pertolinoringenin  Lvs. Acacetin-7-p-glucuso-glucuronide  Lvs. McNudin  Soutellarean, Pertolinoringenin  Lvs. McNudin  Soutellarean, Pertolinoringenin  Lvs. McNudin  Soutellarean, Scutellarean, Acacetin  Angenin, Inteclin  Angenin, Inteclin  Angenin, Acacetin  Angenin  Angenin, Acacetin  Angenin  Angeni	Citherectium subserveium  1.7.3 - alu0me-j,6,4 - tri-OH flavone  Clerodendrum indicum  1.8. Any genin, Accetin  1.8. Any genin, Accetin  C. incritolium  C. incritolium  C. incritolium  C. phicutonium  D. Accetin-7-phicutonium  D. Accetin-7-	Callicarpa_macrophylla	Lvs.	5,4'-d1-OF, 3,7,3'-tri-Ome flavone	Chaudhary, et al. (1978).
infortune indicum  l's. Gincur, oncides of inspidulin, Soutellarenn  lis. Augenin, Acacetun  livs. 7-c glucu: Onde of spigenin and soutellarenn  livs. Soutellare: 1-4: -grabinoside, soutellarein  Livs. Soutellare: 1-4: -glucuso-glucuronide  6-OH inteolin, Gaffeto and Ferulit acid.  Livs. Acacetin-7-4-glucoso (1-2)4-glucuronide  1.vs. Lutsolin  2.ss. Lutsolin  3.ss. Lutsolin  4.ss. Lutsolin  5.ss. Lutsolin  5.ss. Lutsolin  6.ss. Lutsolin	Hys. Hispidilin, Scutellarenn  Incortunatum			7,3'-d10me-j,6,4'-tr1-OH flavone	
infortumetum  1°3. Gincur., and sea agivones.  73. Apigenin, Accetin  1/8. 7-c gincur.onade of spisenin and soutellarein  1/8. 7-c gincur.onade of spisenin and soutellarein  2/8. Soutellarein.4-arebinoside, soutellarein  2/8. Soutellarein.4-gincuso-gincurningenin  1/8. Soutellarein.6-gincuso-gincurningenin  1/8. Accetin-7-p-gincuso-gincurningenin  1/8. Accetin-7-p-gincuso-gincurningenin  1/8. Accetin-7-p-gincuso-gincurningenin  1/8. Accetin-7-p-gincuso-gincurningenin  2/8. Accetin-7-p-gincuso-gincurningenin  2/8. Actedina introlin  2/8. Actedina introl	infortune tum  1'9. Out no free agivones.  73. Augment, scaetta  1'vs. 7-c glucu-conde of apidenin and soutellarein  1'vs. 5cutellare; 1-4'-arabinoside, scutellarein  1'vs. 5cutellare; 1-4'-arabinoside, scutellarein  1'vs. 5cutellare; 1-4'-arabinoside, scutellarein  1'vs. 5cutellare; 1-4'-arabinoside, scutellarein  1'vs. 6-OH inteolin, Caffeto and Ferulic acid.  1'vs. Acacetin-7-4-glucuso-glucuronide  6-OH inteolin, Caffeto and Ferulic acid.  1'vs. Acacetin-7-4-glucuso-glucuronide  1'vs. Acacetin-7-4-glucuso-glucuronide  1'vs. Acacetin-7-4-glucuso-glucuronide  1'vs. Acacetin-7-4-glucuso-glucuronide  1'vs. Acacetin-7-4-glucuso-glucuronide, lutso.  2's glucu-onid., luteolin  2's floctonide, Apigenin, luteolin, Acacetin, Acacetin, Acacetin, Acacetin, Luteolin, Acacetin, Acacetic, Acacetin, Acacetin, Acacetic, Acacetin, Acacetic, Acacetin, Acacetin, Acacetic, Acacetic, Acacetin, Acacetic, Acacetin, Acacetic, Acacetin, Acacetic, Acacetic	Clerodendrum indicum	Lvs.	Hispidulin, Scutellarein	Maj 1
nerne lvs. Apigenin, Acacetin livs. 7-c glucu-onide of apigenin and soutellarein nerifolium lvs. Scutellarein-44 -arabinoside, scutellarein hacacetin, fitspidulin.  Lvs. Scutellarein and Pectolinoringenin lvs. Scutellarein, Caffete and Ferulic acid.  trichotomum lvs. Acacetin-7-p-glucoso (1-2)p-glucundide lvs. Mglvidin Scutellarein, Pectolinoringanin lvs. Mglvidin Scutellarein, Pectolinoringanin lvs. Mglvidin Scutellarein, Pectolinoringanin lvs. Lutsolin, Lateolin, Lutsolin, Lutsol	inerme  Ivs. 7-c glucu.conde of apigenin and soutellarein nerifolium  Ivs. 8cutellarein-4"-arabinoside, scutellarein nacectin, Hispidulin.  Lvs. Scutellarein, Hispidulin.  Lvs. Scutellarein and Pectolinoringenin  Lvs. Acacetin-7-p-glucusu-glucuronide  6-oH inteolin, Caffeto and Ferulic acid.  Lvs. Acacetin-7-p-glucusu-glucuronide  Lvs. Melvidin  Soutellarein, Pectolinoringenin  Lvs. Lutsolin  Angenin, inteolin, inteolin, inteolin, inteolin, daeneteagetin, apigenin, Lutsolin, daeneteagetin, apigenin, Lutsolinoside, Api-7-glucuronide, Lutso-7-glucuronide, Lutso-7-glucur		t.	Glucurunoides of Alspidulin, Soutellarein but no free aglycones.	S.S. andR.Neir
inerme  lvs. 7-c glucu-conde of apisenth and soutellarein nerifolium  cacetin, fispidulin.  Lvs. Scute-larein and Pectolinoringenin  Lvs. Scute-larein and Pectolinoringenin  Lvs. Acacetin-7-p-glucuso-glucuronide  6-OH inteolin, Caffetc and Ferulic acid,  Lvs. Acacetin-7-p-glucoso (1-2)p-glucunonide  Lvs. Acacetin-7-p-glucoso  Lvs. Acacetin-7-p-glucoso  Lvs. Acacetin-7-p-glucoso  Lvs. Acacetin-7-p-glucoso  Respisation  Apigenin, Loteolin, Apigenin, Luteolin,  Apigenin, Loteolin, Apigenin, Luteolin,  Acacetic Api-7-glucoronide, Luteo-7-  Respisation  Apigenin, Loteolin, Apigenin, Luteolin,  Acacetic Api-7-glucoronide, Luteo-7-  Respisation Apigenin, Loteolin, Apigenin, Luteo-7-  Respisation  Apigenin, Apigenin, Luteolin,  Acacetic Api-7-glucoronide, Luteo-7-  Respisation  Apigenin, Apigenin, Luteolin,  Acacetic Api-7-glucoronide, Luteo-7-  Respisation  Apigenin, Apigenin, Luteolin,  Acacetin-7-p-glucoronide, Luteo-7-  Respisation  Apigenin, Apigenin, Apigenin, Luteo-7-  Respisation  Apigenin, Apigenin, Apigenin, Apigenin, Apigenin,  Acacetin-7-p-glucoronide, Luteo-7-  Respisation  Apigenin, Apigenin	nerifolium ivs. Scutellare:1-4'-arebinoside, scutellarein and soutellarein cacetum, filspidulin.  Lvs. Scutellarein-4'-arebinoside, scutellarein cacetum, filspidulin.  Lvs. Scutellarein and Pertolinoringenin  Lvs. Acacetum-7-B-glucuso-glucuronide  Lvs. McPardin  Lvs. McPardin  Lvs. McPardin  Lvs. McPardin  Scutellarein, Pectolinoringenin  Lvs. McPardin  Scutellarein, Surrotin.  Guercetagetin, Apigenin, inteolin,  Apigenin, Lutsolin  Litsolin  Fls. Lutsolin  Apigenin, Lutsolin, Apigenin, Anteolin,  Apigenin, Lutsolin  Fls. Apigenin, Lutsolin,  Anteroeragetin, Apigenin, Lutsolin,  Anteroeragetin, Apigenin, Lutsolin,  Apigenin, Lutsolin,  Apigenin, Lutsolin,  Apigenin, Lutsolin,  Anteroeragetin, Apigenin,  Anteroeragetin, Apigenin,  Anteroeragetin, Apigenin,  Anteroeragetin,  Anteroer		Fls.	Apigenin, Acacetin	Sinha et al. (1981).
institution  Insti	interitolium  Live. Scutellare. 1-4. erabinoside, soutellarein  Entratum  Live. Scutellarein and Pectolinoringenin  Live. Acacetin-7-p-glucuso-glucuronide  6-0H inteolin, Caffeic and Ferulic acid,  Live. Melvidin  Soutellarein, Pectolinoringenin  Live. Inteolin  Live. Linteolin  Live. Linteolin	C. merme	L'vs.	7-c gluco.conde of apigenin and soutellarein	Subramanian, S.S., A.G.R. Mair and T.N.C. Vedan cham (1973).
Lvs. Soute leren and Pectolinoringenin	Ins. Soute leron and Pectolnoringenin  Lvs. Acacetin-/-glucuso-glucuronide  G-OH Luteolin, Caffeto and Ferulic acid.  Lvs. Acacetin-7-p-glucoso (1-2)p-glucuronide  Lvs. Acacetin-7-p-glucoso (1-2)p-glucuronide  Lvs. Melvadin  Soutellaren, Pectolinoringenin  Lvs. Luteolin  Apigenin, Introlin, Quarcetin.  Guareetagetin, Introlin, Quarcetin.  gluculonia,  Luteolin  Fls/ Apigenin, Luteolin, Waercetin.  Guareetagetin, Apigenin, Inteolin,  Apigenin, Luteolin, Waercetin.  Shemplarci, Maemp-2-rutinoside, Api-7-glucuronide, Luteolin,  Angenin, Luteolin,  Angenin, Luteolin, Marcetin.  Shemplarci, Maemp-2-rutinoside, Api-7-rutinoside, Api-7-rutin		£V5.	Scutellare 17-41-arabinoside, scutellarein Acacetin, Hispidulin.	Subramanian, S.S. and A.G.R.Nair (1972).
trichotomum  Lvs. Acacetin-/-glucuso-glucuronide  G-OH inteolin, Caffete and Ferulic acid.  Lvs. Acacetin-7-p-glucoso (1-2)p-glucunonide  Lvs. Mglvidin  Soutellaren, Pectolinoringenin  Lvs. Luteolin  Apigenin, Jutrolin, Jutrolin,  Guercetagetin, Apigenin, Luteolin,  Lutinoside, Api-7-glucuronide, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin, Apigenin, Luteolin,  Apigenin, Luteolin, Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Apigenin, Apigenin,  Apigenin, Apigenin, Luteolin,  Apigenin, Apigenin, Luteolin,  Apigenin, Apigenin, Apigenin,  Apigenin, Apigenin, Apigenin, Apigenin,  Apigenin, Apigenin, Apigenin, Apigenin, Apigenin, Apigenin,  Ap	Lvs.   Acacetin-/-glucuso-glucuronide	C. phlendis	Lvs.	Scute larein and Pectolinoringenin	Subramanian, S.S. and A.G.R.Mair (1972).
trichotomum  Lvs. Acacetin-7-p-glucoso (1-2)p-glucuronide  Lvs. Melvidin  Soutellarein, Pectolinoringenin  Lvs. Luteolin  Angenin, Jatrolin, Jarcetin.  Quercetagetin, Millerolin,  Lampfarol, Kasmp-3-rutinoside, Luteo.  gluculonide,  Luteolin  Fig. Luteolin, Apigenin, Luteolin,  Kaempfarol, Kasmp-3-rutinoside, Api-3-  rutinoside, Api-7-glucuronide, Luteolin,  Kaempfarol, Kasmp-7-rutinoside, Api-3-  rutinoside, Api-7-glucuronide, Luteo-7-  glucuronide, Carotenoide, Luteo-7-	trichotomum  Lvs. Acacetin-7-p-glucoso (1-2)p-glucuronide  Lvs. Wrlvidin Soutellaren, Pectolinoringenin  Lvc. Luteolin  Angeni, Introlin, Introlin, Introlin,  "Angeni, Introlin, Introlin,  "Angeni, Introlin,  "Angenin, Luteolin,  "Frs. Quercetagetin, Apigenin, Luteolin,  "Angenin, Luteolin,  "Angenin, Inteolin,  "Angenin, Apigenin, Luteolin,  "Angenin, Apigenin, Luteolin,  "Angenin, Apigenin, Luteolin,  "Angenin, Angenin, Apigenin,  "Angenin, Apigenin, Luteolin,  "Angenin, Apigenin, Apigenin,  "Angenin,  "Angenin, Apigenin,  "Angenin,  "Angenin, Apigenin,  "Angenin,  "		Lvs.	úcacetin-/-glucuso-glucuronide	Okigawa, et al. (1971).
<pre>Live. Acacetin-7-p-glucoso (1-2)p-glucuronide</pre>	trichotomum  Lvs. Mclvidin  Soutellarein, Pectolinoringenin  Line arboree  Lvc. Luteolin  Apigenin, Introlin, Sucretin.  Guercetagetin, Apigenin, Luteolin,  Lutinoside, Api-7-glucuronide, Luteo.  Lutinoside, Api-7-glucuronide, Luteo.  Lutinoside, Api-7-glucuronide, Luteolin,  Apigenin, Luteolin, Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigenin, Luteolin,  Apigen		-		Nair, A.G.R., T.W.C. Vedantham and Kanaabiran, B. (1979).
Scutellarean, Pectolinoringenin   Scutellarean, Pectolinoringenin   Scutellarean, Pectolinoringenin   Lvs.   Luteolin   Angenin, Luteolin,   Raempferch,	Soutellaren, Pectolinoringenin   Soutellaren, Pectolinoringenin	C. trichotomum	Lvs.	Acacetin-7- $oldsymbol{ ext{-}B} ext{-}$ glucoso (1-2) $oldsymbol{ ext{F}} ext{-}$ glucuronide	Okigawa, et al. (1971).
LVE. Luteolin  Apigenin, Introlin, Charcetin.  Guercetagetin, Apigenin, Inteolin,  Lutinoside, Api-7-glucuronide, Luteo.  glucuronide, Api-7-glucuronide, Luteo.  Fis. Lutiolin,  Apigenin, Luteolin,  Kaempferel, Kaemp-7-rutinoside, Api-7-glucuronide, Luteo-7-glucuronide, Luteo-7-glucuronide, Luteo-7-glucuronide, Luteo-7-glucuronide, Carotenoids.	Luteolin Apigenin, Jutrolin, Quercetin.  Guarcetagetin, Apigenin, inteolin, Apigenin, Apigenin, inteolin, Apigenin, Luteolin, Luteolin, Apigenin, Luteolin, Luteolin, Apigenin, Luteolin, Luteolin, Luteolin, Luteolin, Luteolin,	Duranta repens	Lvs.	reın,	Forsynth, W.G.C. and N.W.Sımmonds(1954). Subramanian, S.S. and A.G.R.Nair (1972).
Apigenin, Jucrolin, Jucrolin, Fao, Guercetagetin, Fingenin, Luteolin, Hair, Laryplarol, Kasmp-3-rutinoside, Luteo. —  glucu-onide, Api-7-glucuronide, Luteo. —  Fis, Luteolin, Apigenin, Luteolin, Apigenin, Luteolin, Apigenin, Luteolin, Kasmp-3-rutinoside, Api-7-glucuronide, Luteo-7-glucuronide, Carotenoids.	Apigenin, Jurolin, Lateolin, Mari-3-  Guarcetagetin, Mission, Mission, Mair, Mair, Mair, Maringlae, Mpi-7-glucuronide, Lutso, -  glucuronide, Api-7-glucuronide, Lutso, -  Els/ Lutsolin, Muercetin, Mission, Marieolin, Mar		Lve.		Rao, et al. (1967).
assetica  Lvs/ Lutsolin  Pls/ Lutsolin, Quercetin.  Frs. Quercetagetin, Apigenin, Luteolin, Kaenpfercl, Kaemp-3-rutinoside, Api-3-  rutinoside, Api-7-glucuronide, Luteo-7- glucuronide, Carotenoids.	assetica  [New Large Lar	2000		, ,	
Instance in the color wherether in the color wherether in the color where the color where the color with the color	Instantant intection, wheretin.  Frs. Auercetagetin, Apigenin, Luteclin, Kaempferel, Kaemp-3-rutinoside, Api-3- rutinoside, Api-7-glucuronide, Lutec-7- glucuronide, Cerotenoids.	. ~		-	
,	,	G. asy ation	Lvs/ Frs.	Lutsolin Apigenin, Luteolin, Quercetin. Quercetagetin, Apigenin, Luteolin, Kaempferch, Kaemp-3-rutinoside, Api-3-rutinoside, Luteo-7-	
				glucuronide, Carotenoids.	Contd

Table : 12. Contd.

Plant name	Purt	Compound reported	Author
Livvia nodiflora	Lvs.	Nodiflorin, A and B	Joshi, E.C. and D.S. Bakuni (1959).
, .	, S	Nodiflorin, a and B	Toshi, B.C. (1966).
		Nodifloretin-A (5,6,7,4'-terra-OH,3' Ome flavone)	Barua, et al. (1969),
		Lippiflorin A,B; 6-0H-Luteolin 6-0me luteolin (nepatin)	Nair, et al. (1973).
Stachyturoheta indica	Lws.	Scutej larein	Subramanian, S.S., A.G.R.Wair and T.N.C. Vedantham (1974).
· ·	-	Choline, chlorogenic acid, 6-0H luteolin, apigenin, glucuronide.	Duret, S.H., Jacquenin and E.A. Paris (1976).
Vitex sp. (V. sanus-castus, V. trifolia, V. negundo	Lvs. or Wood	Castioin (3',5'-di-OH,3,4'-6,7, tetra-Ome flavone), luteolin 7-glucoside, Orientin er l'nomocrientin.	Mansel, et al. (1965).
V. negundo	lvs.	5-0H, 5, 6, 7, 3', 4' -penta-Ome-flavone.  n-CH Pengole sold, 3, 4 di-Oh bengole sold	banerjee, <u>et al</u> . (1969). Joshi, V., J.x. Merchant and U. Nadkarni (1974).
	t. cari	Vanillic, p-C' benzoic acid, Luteolin. Leocoanthocyanin 3,6,7,3,4'-penta Ome 5-o glucopyran(3yl r'amnoside, vitexin, 4'-Ome myricetin-3-o galactopyranoside.	Codanda Rao, E.V.Rao and D.V.Rao(1977) Subramanion, F.M. and G.S.Misra (1978) Hisra, G.S. ahu P.M. Ubramanian (196x).
V. Incens	Ht, wood		Perkin, A.G. (1900). Rao, C.S. and V. Yenkateswarulu (136 ).
	-	Vıtexin	Rao, C.S. and V. Venketeswarulu (1962).
V. leucoxylon		Vıtexin	Rao, P.S. (1965).

Table: 12. Contd.

	Don't	Description of Description	204+ **
	י קקד נ	nas roda i nimodino.	At that
LEGNANS	,		東北の大学の
Gnelina_arborea	Ht, wood	Gmelinol, n-octanosonol.	Joshi, K.C. and L.B. Singh (1970).
i E		Arboreol	Govindachari, T.R., P.C.Parthasarthy
	•		all little Depart (12/2/2)
		. Gmelinol	Desal et al. (1973).
		Arborecl, isoarborecl, Gmelenone.	Anjaneyulu, <u>et al</u> . (1975).
		6' bromoisoarboreol, Gummadiol.	angeneyulu, et al. (1977).
Quinones			
CVIC ennia tomentosa		Layerholl	Sournot, K. (1914).
lectona grandis	Root	Lebachol §	Joshi, K.C., F. Singh and
		Denyd: otectol (Pet. ether ext.)	R.T. Pardasanı (1977).
ر			
	, ,	<pre>p-lapachone Dehyuro- &lt;-lapachone and Tectol { (ethanol ext.)</pre>	
	Lvs.	1,4,5.8-tetra-Hydroxy-2-isopentadienyl	
	· .	anthraquinoné	T.A. Sheshadri (1965).
		Sunnone $A_{6}$ , $A_{7}$ (o-quinoid structure belonging to nuphthalene series).	Sandermann, 4, and M.H. Simatupeng (1955).
	Stem trssue oultre	2,5,d1 OF-1-Ome-3-methyl anthragoinone	uhruwa, <u>et al</u> . (1973).
ALKALOIDS			
. Clerodendrum inerme	Lvs,	A bitter principle, elkaloidal in nature.	Apdul Alm (1971).
C. serratum	300t	Alkeloidal matter absent	danerjee, et al. (1969).
Duranta repens	Fruit	Pyridine alkaloid	Yousef, et al. (1973).
Gmelina arrorea	Lvs.	.lkal.ıds present	Ehattacherjee, A.K. & A.K.Das (1969).
Verbens officinalis	Aeria	Alka-olds present	Ismellov, M.N. (1967).
Vitex negundo	INS.	Miscriniânne alkaloiâ	Basu and Singh (1944). Joshi, K.C., J.R. Werchant & U. Nedkarny (1974)

Contd...

Trable : 12. Contd.

Plant name	1 art	Compound reported	Author.
STEROIDAL SAPONINS			
Avicennia officina is	Lvs/	p-Sirosterol, lupsol, lupenone	Subremennan, S.S. and T.W.C. Vedentham (1574);
Callicarna arrorea	Lvs.	β-Sitosterol	Sen, M. and U. Sarkar (1978).
Clerodendron informatum	Lvs.	Clerodin	Banerjee, H.N. (1937). Chandbary, D.N. and P.C. Dutt (1051).
	Lvs/Root	r p-91 tost_rol	Sen, A.B. and S.P. Singh (1964).
,	Root	<pre>p-Sitosterol</pre>	Barua, A.K., P. Sanyal and P.Chakrabarti (1967).
	Ivs.	Clerodin	Subramanian, S.S. and A.G.M. Nair (1973).
Clerodendrum paniculatum	ನಿಂದಕ	β-amyrın, B-Sıtostarol	Joshi, K.C., P. Singr andlka Mehre (1979).
C. 1 Tomidis	Sten	B-litosterol	Gupt., R.K., Suresh Chandra and V. Kahadevan (1967).
C. serratum	Root	r-3itosterol	bancrjee, <u>st sl</u> . (1969).
Dure 1 ta repens	Frus t	β-5_tosterol	Kapil, R.S. (1960).
Omelina arborea	Lvs.	β-31 tostarol. β-11 tosterol	Reo, b.V. andV. Reo (1970). Joshi,C., L. Prekash s.id b.b.Singh (1971). Desai, et al. (1975).
<u>Lantona comara</u>	Seeds	λ-εωγιια, β-5itosverol β-Sitosverol	Abmed, et al. (1972).
Stachyrarpheta indica	Lvs.	Stigmaeterol	Walr, A.G.F. and T.N.C. Vedentham (1974).
Tectona grandis	Bark	Sagonin yr sent	thattachergee, A.K. and A.K.Das (1969).
Vitex negundo		β-s' tosterol	Josni, et al. (1974).
	. sark	B-Sitosterol .	Kodenda Rao, et al. (1977).
V. trifolium	Lvs.	<b>p</b> -Sitesterol	Vedantham, T.M.C. and S.S.Subramanian (1976).
			Contd

Table: 12. Contd.

			And the second of the second o
IRIDOIDS			
Citharexvlum sp.		Lamiide. Durantoside	Milz, S. and H. Rimpler (1979).
Durenta plumerei	L's/St/ Fruit	Laminde, Durantoside-1,2,-3. Lamilde, Durantoside	Rimpler, H. and T. Helmut (1974). Rao, B.Ch., T.N.Rao and E.K.S. Vijaykumar (1978).
Lantana camara	Lvs/St.	Theveside (Iridold glucoside)	Ford, C.W. and W.R. Bendall (1980).
Premna latifolia		7-deoxyloganıc acid, genıposidic acid	Rao, B.Ch., E.K.S. Vijaykumar and K.V. Vijayalaxmı (1981).
Etachytarpheta induca	A.r.al	Ipolamide .	Tantisewie, 5. and Ottosticher (1975).
S. mutabilis	par us	Ipolamıde	Garnier, J. (1979).
Verbena pulchella		Pulchellyside II.	ililz, S. en.1 4. Rimpler (1978)
Verbena su.		Verbenallune, Hastatoside, Griselinoside, Lanide, Polohelloside I & II	wilz, S. and H. Rimiler (1979).
· ds · \hat{\lambda}		Cornin, Hastatosiue, Griselinoside, Laminde, Pulchilloside I and II.	Lemtoft, <u>et al</u> . (1979).
V. hastata		Verbenalin, Hastatoside	Rimpler, H. and B. Schafer (1979).
Vitex meganotamics:		Ágnoside	Rimpler, E. (1972).

not much explored. There are random reports of flavonoids and other phenols from individual plants. These isolated reports are indicative of our meagre knowledge about the phenolic chemistry of this family. Not a single plant has been worked out thoroughly and systematically for phenolics. In the present work, 35 plants belonging to 14 genera have been systematically screened for the various phenolics. The leaf phenolics taken into consideration here include the flavones, flavonols, glycoflavones, leucoanthocyanins, quinones and phenolic acids. Therefore, in the present report an attempt is made to understand the interrelationships and phylogeny of Verbenaceae using reliable chemical markers. Data obtained from other disciplines of Botany are also properly weighed along with the chemical data for a proper appraisal of the present classifications of the family and to discern the evolutionary trends within this large taxon.

#### 3:4 Materials and Methods

Most of the screening was done with fresh materials collected from gardens around Baroda. Supply of materials also came from the Botanical gardens at Waghai (Dangs), Aurangabad, Pune and Trivandrum. Premna resinosa was made available by Dr. K.S.S. Rao from the Kutch region. Avicennia

was sent from Bulsar by Dr. P.G. More. The plant materials were properly dried, preserved and deposited in the Herbarium, Botany Department, The M.S. University of Baroda, Baroda (BARO).

The procedures followed in extraction, isolation, and identification of the various phenolics are the same as those described in Chapter 2.

#### 3:5 Results

The distribution of flavonoid compounds (such as Flavones, Flavonols, Glycoflavones, Leucoanthocyanins), Phenolic acids, Alkaloids, Iridoids, Saponins, Tannins and Quinones from leaves of Verbenaceae members is presented in Tables 13, 14 and 15.

In the present work 35 members belonging to 3 tribes and 14 genera were studied using chemical markers like flavonoids and phenolic acids. 31 members contain one or the other type of flavonoids. 4 members are devoid of any flavonoid. A total of 30 o-glycosides and c-glycosides are detected, of which 19 are of flavones, 7 of flavonois and 4 c-glycosides.

Flavones are found singly in 19 members. Flavones and Flavonols occur together in 8 members. Flavones along with c-glycosides occur in 3 members. While Flavones/Flavonols/

Table : 13. DISTRIBUTION OF FLAVONOIDS IN THE VERBENACEAE.

No. MAME OF THE FLANT		2 7 + 2 4	17 19 19 50 51 55 52 54	
TRIBE VERBENEAE 1. Citharexylum subserratum Sw.	+ +			+
2. Duranta rapen. Tinc.	† †	+		
3. Lantana camara var. acu 'ata (L.) isc denke	+ +	•	+	
	# +	1	Application of the state of the	To the state of th
5. var. mista Bailey	+		2	•
6. var. inivea Bailey	+ +			•
7. L. salvifolia Jaco.	+	+	+	
8. Petres volubilis Jena.		ŕ		
9. Phyla nodiflora (Lidn.) Greene		.+	, . +	
10. Stachytarpheta india Vahl.	+	•	÷	
11. S. mutabilis Vahl	+		+	
12. Verbena-venosa Gill; & Hook.	+			1
13. 7. bipinnatifida M.t.	+		÷	+
14. V. tenuisecta Bric.	+			
TRIBE ' / - COE4E				
15. Clerodendrum sculestum (Jann.) Grassb.				
16. C. fragrans Vent.	+			
17. C. indicum (Linn.) Evntse	*	+		+
15. C. inerwe (Lina.) Gaertn.	+			
19. C. infortunatum Gaertn.	-			
2(. C. macrosiphon Hook. f.	÷			
21. C. , ultiflorum (Burm.f.) O. Ktze.	ŀ	+	-	-
22. C. paniculatum inn.	+ .	•	-	
23. Clerodendrom slobchenthus R. Br.	+ +	-	٠	
•				

Table: 13. Contd..

Sr. NAME OF THE PLANT ,	1234567891	10 11 12 13 14 15 16 17 18 19 20 21 2	22 23 24 25 26 27 28 29 30
25. C. speciosum Fart.	+ + *+		
27. Gmellina arhorea Rox'. 28. Grasia do allam. 29. Graniliopensis Cha.	+ + + 1	+ +	+
	+	, + +	+
33. Vitex negundo L. 34. V. altissima Lion. 4. TRIBE SYSTEMMERE	+	+	
35. Avicennia manina (Forsk.) Vierh. Syn. A. alti aurt. non Blume		+	+
1. Apigenin 2. 4'-Ome Apigenin 2. 7'-Ome Apigenin 4. 'altellaren 5. 4!-Ome Scutellarein 6. 5-Ome 7. 6-Ome 10. 7,4'-di Ome Scutellarein 10. 7,4'-di Ome Scutellarein	11. Luteolin 12. 5'-Ome Luteolin 13. 4'-Ome " 14. 6-Ome " 15. 7',4'-dı Ome Luteolin 15. 7,4'-dı Ome Luteolin 17. 6-O''-Luteolin 18. 6-O! 19. 6-O'',2'',4'-qıO''E Luteolin 19. 6-O'',2'',4'-qıO''E Luteolin 19. 6-O'',2'',4''-qıO''E Luteolin 19. 6-O''',2'',4''-qıO''E Luteolin 19. 6-O''',2'',4''-qıO''E Luteolin	Flavonols  20. Kaempferol  21. 6-014! -Ome Kaempferol  22. Quercetin  23. 3! -Ome Quercetin  24. 4! -Om;  25. 3', 4! -\in Ome "  26. 6-01. Quercetin (Quercetegetin)	Glycoflavones 27. Vitaxin 28. 7-One vitaxin 29. G.F.besed on 7-One Apiganin 30. b-Ore Orientia.

Glycoflavones together are present in only one member belonging to tribe Avicennieae.

14 members belonging to tribe Verbeneae were analysed for flavones/-ols. The results clearly indicate dominance of flavones. All the 14 members show presence of one or the other type of flavones. 8 members show exclusively flavone type of compounds. They are <u>Duranta repens</u>, <u>Lantana camara var. aculeata</u>, <u>Phyla nodiflora and Verbena tenuisecta</u>. The other 5 members possess both flavones and flavonols, as encountered in case of <u>Citharexylum subserratum</u>, <u>Lantana salvifolia</u> (= <u>L.indica</u>), <u>Stachytarpheta indica</u>, <u>Stachytarpheta mutabilis</u> and <u>Verbena bipinnatifida</u>. <u>Verbena venosa</u> is the only member from this tribe where flavones along with glycoflavones are detected.

The different flavones identified in this tribe are mostly methylated or hydroxylated derivatives of Apigenin and Luteolin. Apigenin itself is detected in 6 members belonging to this tribe. They are Citharexylum subserratum, Duranta repens, Lantana camara var. aculeata, var. nivea, Phyla nodiflora and Verbena tenuisecta. The aglycone part of apigenin-glucuronide as reported earlier from Stachytarpheta indica (Duret et al., 1976) has been found to be absent. However, Stachytarpheta mutabilis shows presence of 4-methoxy apigenin (Acacetin). Acacetin is also detected in

<u>Duranta repens, Lantana camara var. aculeata, var. hybrida, var. mista, var. nivea, Stachytarpheta mutabilis</u> and <u>Verbena tenuisecta.</u>

7-methoxy Apigenin, the other monomethyl derivative of apigenin is found in only two representatives of this tribe - Citharexylum subserratum and Verbena venosa.

Scutellarein (6-OH, apigenin) as reported formerly by Subramanian and Nair (1972) from <u>Duranta repens</u> and by Subramanian, <u>et al</u>. (1974) from <u>Stachytarpheta indica</u> is found to co-occur with 4'-Ome scutellarein in leaf extract.

<u>Lantana camara var. hybrida</u> also shows presence of scutellarein. The other two varieties viz. var. <u>mista</u> and var. <u>nivea</u> show presence of 4'-Ome scutellarein. The occurrence of 7,3'-di-Ome, 5,6,4' tri-OH flavone (7,3'-di-Ome Scutellarein) as reported from <u>Citharexylum subserratum</u>

Subramanian <u>et al</u>., (1976) could not be detected during the present flavonoid screening. However, it shows presence of another scutellarein derivative - identified as 6-Ome scutellarein (Hispidulin). 6-Ome scutellarein is also detected in taxa like <u>Lantana salvifolia</u> and is the only flavone in Verbena bipinnatifida.

From the dimethoxylated derivatives of scutellarein, 6,4'-di-Ome scutellarein (Pectolinaringenin) was detected in

Lantana salvifolia and Stachytarpheta mutabilis. But, found to be absent in <u>Duranta repens</u> contrary to an earlier report by Subramanian and Nair (1972).

3'-Ome Luteolin is found only in <u>Duranta repens</u>. The other monomethyl derivative of luteolin such as 6-Ome luteolin (Nepatin) and 6-OH luteolin reported by Nair <u>et al</u>. (1973) from <u>Phyla nodiflora</u> are also detected from the same plant. 6-Ome Luteolin is also found to be present in taxa like <u>Lantana salvifolia</u> and <u>Lantana camara var</u>. <u>hybrida</u>. In addition to these two reported compounds from <u>Phyla</u> nodiflora, 7,4'-di-Ome luteolin is also detected from the same plant.

The earlier report of occurrence of 6-OH luteolin from Stachytarpheta indica Duret, et al. (1976), is confirmed in the present chemical findings. 6-OH, 3', 4'-di-Ome Luteolin is also detected from the same plant.

The 6 flavonol compounds detected in this tribe include 6-OH quercetin (quercetagetin) from <u>Citharexylum subserratum</u>; Kaempferol from <u>Stachytarpheta mutabilis</u>, 6-OH, 4 -Ome Kaempferol from <u>S. indica</u>, Quercetin from <u>Lantana salvifolia</u> and Stachytarpheta mutabilis, 3'-Ome quercetin from <u>Lantana salvifolia</u> and <u>Verbena bipinnatifida</u> and 4'-Ome quercetin only from Verbena bipinnatifida.

20 members of the second tribe Viticeae have been screened. 16 of them show presence of flavones/flavonols. Of these 16, 11 plants are with flavones only, 3 with both flavones and flavonols but no glycoflavone and 2 with flavone and glycoflavone. Thus in this tribe also, flavones are detected in majority of the members. The incidence of flavones, however, is comparatively more in tribe Verbeneae. Flavonols are of relatively rare occurrence.

The various taxa which possess only flavones are Clerodendrum fragrans, C. inerme, C. macrosiphon, C. multiflorum (= C. phlomidis), C. paniculatum, C. siphonanthus, C. splendens, C. speciosum, Gmelina philippensis, Premna resinosa and Vitex negundo. Gmelina arborea, G. asiatica and Tectona grandis are the only members of tribe which possess both flavones and flavonols. Clerodendrum aculeatum, C. infortunatum, C. thomsoniae and Vitex altissima are devoid of any flavonoid compounds.

In all 15 flavones have been identified from Viticeae members. Apigenin is encountered from two members — C. speciosum and Gmelina arborea. The presence of apigenin and luteolin as reported by Rao and Rao (1970) from G. arborea is confirmed. But, instead of quercetin its another methylated derivative (3 -Ome Quercetin) is detected along with acacetin. The other species G. asiatica shows

presence of acacetin, Kaempferol, Quercetin and its dimethylether (3',4'-diOme quercetin) which is in agreement with the earlier report of Nair and Subramanian (1975) except with one additional flavone - the luteolin, identified during the present study. The third species, G. philippensis shows presence of acacetin only.

Out of the 12 <u>Clerodendrum</u> species screened, 9 contain flavones. The flavonols are completely absent. The 3 species totally lack flavonoid type of compounds. Among the flavones and their derivatives, acacetin has been identified from <u>Clerodendrum fragrans</u>, <u>C. inerme</u>, <u>C. microsiphon</u>, <u>C. paniculatum</u>. 7-Ome Apigenin, 4'-Ome scutellarein and 7,4'-di-Ome scutellarein are detected from <u>C. speciosum</u>. The other Viticeae members with acacetin are <u>Holmskioldia sanguinea</u> and <u>Premna resinosa</u>.

Scutellarein, reported by Subramanian and Nair (1972) from Clerodendrum multiflorum and by Subramanian et al. (1973) from C. inerme is also found to be present, confirming the earlier reports. Scutellarein is also present in Tectona grandis and Vitex negundo.

5-Ome scutellarein is detected only from <u>Clerodendrum</u> <u>siphonanthus</u>. Hispidulin (6-Ome scutellarein) reported by Subramanian and Nair (1973) from <u>C</u>. <u>indicum</u> has also been noted.

Luteolin co-occurs with its 3 -Ome derivative in case of <u>Tectona grandis</u>. Whereas, 3 -Ome luteolin and 3 ,4 -di-Ome luteolin are detected from <u>Vitex negundo</u>. 6-OH Quercetin (Quercetagetin) is detected only from <u>Tectona grandis</u>. <u>Vitex</u> altissima is devoid of flavonoid compounds.

The genus <u>Avicennia</u> of tribe Avicennieae shows presence of a flavone, flavonols and a glycoflavone. The flavone and flavonols are 3'-Ome luteolin and kaempferol and its derivative respectively.

In all 4 different glycoflavones (c-glycosides) have been detected. They are vitexin, 7-Ome vitexin, glycoflavone based on 7-Ome apigenin and 6-Ome orientin. The glycoflavones are almost absent from tribe Verbeneae except, in case of Verbena venusa where it is present in the form of glycoflavone based on 7-Ome apigenin. In the 2 members of Viticeae - Clerodendrum indicum and Holmskioldia sanguinea they are present as vitexin and 6-Ome orientin respectively. Vitexin has been reported uniformly in different parts of genus Vitex (Rao and Venkateswarulu, 1962). Its presence is reported in heartwood of V. altissima (Rao, 1965) and bark of V. negundo (Misra and Subramanian, 1980). However, the present study showed the absence of vitexin in both these species. The only member of Avicennieae screened here shows 7-Ome vitexin.

As in Bignoniaceae, the leucoanthocyanin test in most of the Verbenaceae members has been negative, except in <u>Stachytarpheta mutabilis</u> of Verbeneae and <u>Clerodendrum splendens</u> and <u>Tectona grandis</u> of Viticeae.

15 phenolic acids have been detected. p-Hydroxy benzoic, Gentisic, Syringic, p-Coumaric and Ferulic acids are common to all the three tribes. Tribe Verbeneae and Avicennieae show 100% incidence of p-hydroxy benzoic acid, while Viticeae show only 80%. Similarly Gentisic acid has 78.5, 55, and 100% incidence value in three respective tribes (Table 15).

Vanillic acid is quite frequent in tribe Verbeneae and

Viticeae, however, absent in Avicennieae. Syrigic acid is rather more common in tribe Verbeneae than in Viticeae.

Genus Avicennia also shows presence of Syringic acid.

p-Coumaric acid is widely distributed in tribe Viticeae and in genus Avicennia, whereas O-Coumaric acid is frequent in tribe Verbeneae.

Protocatechuic and Salicylic acids are confined to tribe Viticeae. Salicylic acid is detected in Clerodendrum fragrans, C. infortunatum, C. macrosiphon, C. paniculatum and Gmelina arborea. As against this only one Verbeneae member - Phyla nodiflora shows presence of this phenolic acid. Taxa like Citharexylum subserratum, Lantana salvifolia from tribe Verbeneae and Clerodendrum inerme, C. multiflorum,

Table : 14. The Distribution of Phenolic Acids, Leucoarthocyanins, Alkaloids, Iridoids, Saponins, Tannins and Quinoues in the Verbenaceae.

Sr. N	NAME OF THE FLANT	1 2	5 4	R	2 9	ω	9	10	7	12 13	14	15	16 1	17 18	19	20 2	21 22	23.	
C.A	RIBE VERBENE.E															!	1		
٠.	Citharexylum subserratum Sw.	+	+		7	_			+	+		+		+					
2.	Duranta repens Linn.	+	+		*	+		+	+				í				٠.		٠
K,	Lantana camara var. aculeata (Linn.) Moldenke	+	+		7-	4	+							+	+	i	+		}
4.	var. hybrida Moldenke	+	+		+				+	+		1		+	+	+		+	
5	var. mista Bail y	1	4-			+		+		+	+	+	7	+			+		
. 6	, var. nivea Dell'y	+	-4-		*	+		+	+	+			•	+	+	+		+	
I7.	7. Lantana salviiolia Jacq.	+	+		+				+	۲			4		ł			4.	
8	Petrea volubilis Jacq.	+	t.		+	+		4	4	+			4		+		+	+-	
9	Phyla nodiflora (L'nn.) Greene	+	4-	+	7	+	+		+						+		+		
10.	Stachytarphete migica Vahl	+	.4.			+			+	+			+	+	÷		÷		
11.	S. mutabilis vabl	+	+		+	+		+	-{-				+	+	<del>†</del>		+	"	
12. 3	Verbena yenose Gill & lock.	+			4.	+	+	+		+		+	+		÷	-	+		
13. 1	V. bipinnatifi a Mutt.	+			+		. +	+	÷	+		ተ	-1-		+	+	+		
14. 4	V. tenuisecta Briq	+ .			+		+	-	4	÷		+			+			÷	
Ţ	TRIBE VITICAAE.																		
15.	Clerodendrum avrleatum (Lann.) Griseb.	+			+				+							-		+	
16.	C. fragrans Veit.	+ +			+					+		+			+		+	+	
17. 0	C. Indicum (Linn.) Kuntze	+			+								4		+	+			
18.	C. inerme (Linn.) Gaertn.	+	+		+	+		+	+	+			+		+			+	
19. 0	C. infortunatum Gaertn.	+			+				<del>†</del>	+			+		+		+		
20.	C. macrosiphon Hook, f.	+			÷				+	+							+		
21. [	C. multiflorum (Burn. f.) O. Atze.	+	+		+				+	÷	+	+			+			1-	
22. C	C. <u>pantoulatum</u> Lin <del>a</del> .	4			+		+	+	+	+		+			+			4.	

. Contd.

			-	1	1						- 1	1		
Sr. NAME OF THE FLANT	2 3	4 5	2 0	2	8 9	5	11 12	13	14 15	16 17	18	19 20	21	22. 25
23. Clerodendron siphonanthus R. E.	+		+	+		+	+	+		+		+	·	
24. Clerodendrum splendens Don	+			+			+	+	+	+		+	,	+
25. C. speciosum Hort.	+			, <b>-</b> +			+			+		+	,	4
26C. thomsoriae Balf,				+	i		+			. +				
27. Gmelina arborea Roxb.	+			4	+	+	+	+		+			,	
28. G. asiatica Linn.	+			+		+	+	+		+		÷		+
29. G. philippens s Cham.	+			+			+	+		+		+		±-
50. Holmskioldia sanguinea Retz.	+	4		+			+	+	+	+		+		ı
31. Premna resinosa (Hochst.) & hau.	+			+		+	+			+		-		
32. Tectona grand; s Lann. f.	+				ı	+		,	. +	. +		, <b>-</b>	+	+
33. Vitex negunda L.	+	+		+	+	+		+	٠.	+	+	, +	·	4
34. V. altissime inn. f.	+					+	+	÷	•	+		` +		
TRIBE AVICANTEAE														
35. <u>Avicennia mar.na</u> (Forsk.) Vierh. Syn. A. <u>alba</u> auct. non Blume	+	+			+		+	+				4-	•	+
Phenolic solds														
1. Salycyllo 2. p-OH-benzolc Acid 3. Aentleic 4. Protocatrolulo				1. 2. 2. 4.	p-Coume: Phlorat. Ferulic	p-Coumeric Phloratic Fervlic	O H O		Quinones	น es	22.		praqu zocul hthaq	Anthraquinones Benzoquinones Maphthaquinones
5. 2-Hydroxy-5-methory penzon Acid 6. 2-Hydroxy-6-methory " " " 7. Nanillic 8 Syringic 9. 0-Joumanic				15.	Chlo Leve	Chlorogenic Leucoanthoc Alkaloids Iridoids	Chlorogenic  Levicoanthocyaning  Alkaloids  Tridoids  Fanonins	nins			-			

Table: 15. Percentage of incidence of phenolic acids in the tribe Verbeneae, Viticeae and Avicennieae.

PHENOLIC ACIDS	-	Verbenacea (Tribes)	9
	Verbeneae	Viticeae	Avicennieae
Salicylic	7.14	25 .	, _
p-Hydroxybenzoic	100	80	100
Gentisic	78.57	55	100
Protocatechuic	14.28	<sup>,</sup> 20	100
2-Hydroxy 5-methoxy Benzoic	7.14		
2-Hydroxy 6-methoxy Benzoic	653	5	eres
Vanillic	85.7	90	~
Syringic	64.28	20	100
o-Coumaric	35.7	5	
Melilotic	57.14	45	wite
p-Coumaric	78 <b>.</b> 57	80	100
Phloretic	-	5	-
Ferulic	71.42,	70	100
Sinapic	7.14	5	
Chlorogenic	35.7	30	-

Holmskioldia sanguinea and Vitex negundo from tribe Viticeae show presence of protocatechuic acid and so also the genus Avicennia.

2-0H,5-Ome Benzoic and 2-OH,6-Ome Benzoic acids are identified from Phyla nodiflora and Clerodendrum siphonanthus respectively. Melilotic acid is fairly common in various members of tribe Verbeneae and Viticeae but absent in Avicennieae. The phenolic acid in Gmelina arborea gives colour reaction with spraying reagents. However, the colour fades soon afterwards. This indicates doubtful presence of phloretic acid in the plant. Sinapic acid is detected in Lantana camara var. mista and Clerodendrum multiflorum. Chlorogenic acid is common to Verbeneae (35%) and Viticeae (30%) but absent in Avicennieae.

Thus, from the distribution pattern of various phenolic acids in Verbenaceae, it is clear that a number of phenolic acids are common to all the 3 tribes. However, some of them are common to tribe Verbeneae and Viticeae but absent in tribe Avicennieae. The higher incidence of Gentisic, Vanillic, o-Coumaric, Melilotic in tribe Verbeneae, that of Salicylic, Protocatechuic in Viticeae, and absence of Salicylic, Vanillic, o-Coumaric, Melilotic and Chlorogenic in Avicennieae characterises the 3 tribes.

8 members of the Verbenaceae (7 of Verbeneae and 1 of Viticeae) have been found to possess iridoid compounds. Iridoids are absent in Avicennia. The presence of iridoids from Duranta repens (Rimpler and Helmut, 1974) could not be confirmed in the present study. The positive test for iridoids in case of Lantana camara, Stachytarpheta indica and S. mutabilis agrees with the earlier reports (Ford, et al., 1980; Tantisewie and Ottosticher, 1975 and Gurnier, 1979). The higher incidence of iridoids in the tribe Verbeneae as compared to Viticeae and their complete absence in the Avicennieae have taxonomic significance.

Saponins are present in 27 Verbenaceae taxa. Most of them show moderate amount of saponin (+). As in the Bignoniaceae, Tannins are absent in the Verbenaceae.

32 members show the presence of various Quinones. Tribe Verbeneae has benzoquinones and a higher incidence of anthraquinones. Tribe Viticeae has benzoquinones, naphthaquinones and a lower incidence of anthraquinones. Avicennieae, on the other hand has benzoquinones but no anthraquinones.

Around 62% of the plants gave positive test for alkaloids. However, when analysed chromatographically using BAW as solvent system, they did not show clearcut bands either in UV light or in iodine chamber, nor when sprayed with Dragendroff's reagent.

#### 3:6 Discussion

Of the 8 tribes (Bentham and Hooker), only 3 tribes have been worked out in the present study. The chemical data points to the distinct nature of 3 tribes studied, due to their characteristic compounds. The tribe Verbeneae may be considered as chemically advanced due to occurrence of flavones in all the 14 members screened, absence of leucoanthocyanins and glycoflavones accompanied by the presence of iridoid compounds. Tribe Viticeae also shows presence of flavones and flavonols. Although flavones are detected in majority of viticeae members, yet the percentage incidence of flavones is comparatively lesser. Loss of flavonoid compounds in some of the Clerodendrum species marks the advancement of the tribe. However, the tribe is, on the whole, slightly at a lower evolutionary level than the Verbeneae. The presence of a glycoflavone and absence of iridoids in Avicennia, the only member screened in the third tribe Avicennieae appears to be primitive.

The various representatives of these 3 tribes are arbitrarily grouped by Bentham and Hooker based on morphological similarities. The so-called groups are not designated by any taxonomic category. In the present discussion, more emphasis is laid on tribe Verbeneae and Viticeae. An attempt

is being made to utilise chemical data to substantiate grouping of the various members based on morphological (flower and fruit particularly) criteria.

All the 14 Verbeneae members show presence of flavones. Of these 14 members, 5 also possess flavonol type of compounds.

The first group under Verbeneae includes Lantana, Phyla and Stachytaroheta. The genus Lantana as a whole, can be considered as an advanced member within the tribe Verbeneae due to higher incidence of flavones, iridoids and absence of glycoflavones and leucoanthocyanins.

Lantana camara and its 4 varieties studied in the present investigation show presence of acacetin in the leaf extract uniformly. However, the 4 varieties can be distinguished on other chemical grounds. Lantana camara var. aculeata and nivea, have apigenin as the flavone. But, the presence of 4 -Ome scutellarein along with anthra- and naphthaquinones in var. nivea and that of 6-OH luteolin and benzoquinones in var. aculeata clearly indicates that these two are distinct from each other not only morphologically but chemically as well. Similarly, var. hybrida with Scutellarein, 6-Ome-luteolin, anthra- and naphthaquinone and absence of alkaloids is quite different from var. mista,

which shows presence of 4 -Ome scutellarein, alkaloids and benzoquinones.

The presence of iridoids in all the 4 varieties points to the distinct and advanced chemical feature of Lantana camara; thus characterising itself from the other species - L. salvifolia. The distribution pattern of flavonoids in the two species of Lantana also reveals chemical differences at species level. L. camara and its 4 varieties show apigenin and scutellarein type of flavones. Whereas, L. salvifolia is found to contain scutellarein and luteolin derivatives along with quercetin and its methyl derivatives. The presence of protocatechuic acid and absence of anthraand benzoquinone, iridoids in L. salvifolia also substantiate the species difference.

Phyla nodiflora is characterised by presence of only flavones. Some of them it shares with Lantana.

Stachytarpheta possesses both flavones and flavonols.

The two species of Stachytarpheta show different o-glycosides.

S. indica contains 4 flavones and one flavonol. Whereas,

S. mutabilis shows acacetin and scutellarein derivative as flavones and quercetin and kaempferol as flavonols. The absence of phenolic acids (Vanillic and Melilotic), leuco-

anthocyanins and presence of only benzoquinone characterise

S. indica. On the other hand, S. mutabilis not only possesses all these compounds but has in addition naphthaquinones also.

Thus the arbitrary grouping of Lantana, Phyla and Stachytarpheta based on the ovary and fruit characters is justified on the basis of chemical findings. Phyla is chemically closer to Lantana than to Stachytarpheta.

Genus <u>Verbena</u>, which represents another group in Verbeneae shows flavones, flavonols and glycoflavones and absence of iridoids. The 3 species of <u>Verbena</u> screened, reflect chemical differences in their pattern of flavonoid distribution. The only common feature among these species is the presence of apigenin or its derivative as flavone. This supports their placement under one genus.

V. venosa reveals the presence of both advanced and primitive character in the species. The occurrence of 3'-methoxy quercetin and 3',4'-dimethoxy quercetin in addition to a single flavone - 6-methoxy scutellarein in V. bipinnatifida is suggestive of its primitive nature. V. tenuisecta appears to be an advanced species of the genus. The quinone distribution pattern also shows distinct identity of these species. V. venosa shows presence of a benzoquinone, V. bipinnatifida -

with anthra- and benzoquinones, whereas <u>V</u>. <u>tenuisecta</u> shows only naphthaquinone.

The occurrence of a single flavone (6-hydroxy luteolin) in <u>Petrea volubilis</u> may be considered as an advanced feature leading to the elimination of entire flavonoid system.

The morphological grouping of <u>Citharexylum</u> and <u>Duranta</u> under a group in the Verbeneae is not supported on the basis of present study. Except for Apigenin, the distribution of other flavones is quite different in the two genera. The presence of quercetagetin, iridoids and complete absence of quinones in <u>Citharexylum</u> and absence of flavonols, iridoids and presence of quinones in <u>Duranta</u> indicate their chemical differences. <u>Duranta</u> shows a number of chemical similarities with Lantana rather than Citharexylum.

Tribe Viticeae has been divided into two broad categories, based on the corolla and fruit characters. The first group include members like <u>Tectona</u> with regular corolla and 1-4-seeded pyrene fruit.

Tectona grandis is considered as the most primitive taxon due to retention of 5 fertile stamens in the flowers. The present chemical data also point to its primitiveness due to presence of flavonols, leucoanthocyanins and absence of iridoids. Anatomically however, it is considered an

advanced genus due to presence of ring-porous wood, paratracheal banded parenchyma (Metcalfe and Chalk, 1950; Chawdhary, 1964).

The second group in the Viticeae is characterised by irregular corolla and 4 didynamous stamens. As in tribe Verbeneae, this is divided into unnamed subgroups, mainly on fruit character.

Premna and Vitex are grouped along with Gmelina. All the 3 taxa show presence of flavones and absence of leucoanthocyanins and glycoflavones. Premna and Gmelina have acacetin in common. However, the presence of other flavonoid compounds indicate some amount of dissimilarity between these two genera. The presence of flavonols and absence of iridoids in Gmelina as against presence of iridoids in Vitex reflect the chemical difference. The pollen grains are also distinct in the two genera. Gmelina is exceptional in the tribe Viticeae having 3-zonocolporate type. While, Vitex and Premna have 3-zonocolpate type (Nair and Rehman, 1962).

The 3 species of <u>Gmelina</u> screened in the present study are also found to be chemically distinct. Acacetin is common to all of them but <u>G. arborea</u>, in addition to acacetin, shows presence of apigenin, luteolin and 3 -methoxy quercetin.

While G. asiatica has kaempferol and 3,4 -dimethoxy quercetin. The occurrence of acacetin in all the 3 species indicate the homogeneity of this taxon. G. arborea has advanced type of flavonoid compounds - mainly the flavones. The G. asiatica, due to presence of more flavonols than flavones indicates primitive nature. G. philippensis with single flavone may be considered as an advanced feature leading to the elimination of entire flavonoid system. The distinct nature of these 3 species is also evidenced on palynological grounds (Nair and Rehman, 1962).

The two species of <u>Vitex</u> screened, also appear to be distinctive chemically as well as palynologically. <u>V.negundo</u> possesses flavones, iridoids and prolate pollens. The other species - <u>V. altissima</u> is devoid of such compounds and exhibit subprolate pollen grains.

The other subgroup includes <u>Clerodendrum</u> and <u>Holmskioldia</u> due to presence of 4-pyrene drupe. The flavonoid distribution pattern also shows chemical similarity in the two taxa. However, the presence of glycoflavone in <u>Holmskioldia</u> points to its primitiveness within this subgroup. Palynologically also they have similar type of pollen.

The complete absence of leucoanthocyanins (except in Clerodendrum splendens), the presence of flavones and at

times total absence of flavonoid system mark the advancement of <u>Clerodendrum</u>. The flavones when present, are mainly represented by apigenin and scutellarein derivatives except in <u>Clerodendrum</u> indicum which possesses 6-Ome scutellarein, 4'-Ome luteolin, Vitexin and anthraquinone. Occurrence of benzo- and naphthaquinone in most of the species of <u>Clerodendrum</u> are notable chemical characters. The genus, therefore, shows various levels of evolution represented by those with many flavones, those with a single flavone leading to a loss of flavonoid system in <u>Clerodendrum</u> aculeatum, <u>C. infortunatum</u> and <u>C. thomsonae</u>.

In the present study the monogeneric tribe Avicennieae is represented by <u>Avicennia marina</u> which possesses a single flavone, 2 flavonols and a glycoflavone. It also shows absence of leucoanthocyanins, alkaloids and iridoids. Although this taxon combines primitive and advanced characters, the primitive ones appear to have an edge.

Erdtman (1952) remarked that Avicennieae "probably should be regarded as a family of its own" on the basis of clear cut palynological differences. Such a contention has been supported by Nair and Rehman, 1962; Mukherjee and Chanda, 1973; Saxena, 1973, 1975, 1981; on the basis of the palynological studies of Indian members. Metcalfe and Chalk (1950) also considered the Avicennieae as anomalous in the

Verbenaceae and advocated its separation under a separate family on the basis of wood structures. Embryological studies of Junell (1934), Pal (1951), Rao (1952), Padmanabhan (1960), Maheshwari and Kapil (1966) have also suggested isolation of the Avicennieae from rest of the Verbenaceae.

Although there is an overwhelming evidence in support of separate family Avicenniaceae, the chemical characters presented here are not so different from the rest of the Verbenaceae to warrant a separate family status for the genus. Presently the author would be satisfied to retain Avicennieae within the confines of the family Verbenaceae.

It is highly probable that many of the character deviations that the group displays is a direct consequence of its adaptations to peculiar ecological niche. It is, therefore, felt improper to draw sweeping taxonomic conclusions on the basis of ecologically adaptive characters.

All the 3 tribes - Verbeneae, Viticeae and Avicennieae have distinctive chemical characters. However, the differences are of a minor nature and, therefore, raising these groups to subfamilial level is considered unwarranted.

A brief discussion on the taxonomic position of genus Nyctanthes will not be out of place here. The genus Nyctanthes has changed position at the hands of taxonomists. It has been traditionally grouped under Oleaceae. However, Airy Shaw (1952) on morphological and stant (1952) on anatomical evidences proposed its transfer as a new subfamily under the Verbenaceae. In the latest edition of Engler's syllabus (ed. Melchior, 1964), Nyctanthes is grouped under the Verbenaceae. Embryological investigations (Kapil and Vani, 1966), trichome studies (Inamdar, 1967) and anatomical studies on gynoecium (Kshetrapal and Tiagi, 1970) however, do not support the transfer of Nyctanthes from Oleaceae to Verbenaceae. Daniel and Sabnis (1981, in press) have proposed placement of Nyctanthes in the Oleaceae based on chemical characters and numerical analysis.

Chemically <u>Nyctanthes</u> is closer to subfamily Jasminoideae of Oleaceae. It has a high frequency of flavonol and absence of glycoflavones. The Verbenaceae on the other hand, is characterised by both flavonols and flavones (Daniel and Sabnis, 1979).

The presence of iridoids is considered an advanced character (Jensen et al., 1975). Out of the 35 plants screened under the present study, 7 of Verbeneae and 1 of Viticeae show presence of iridoids. This suggests an advanced nature of the tribe Verbeneae within the family Verbenaceae.